Hypochlorous Acid

1	Identification of	of Peti	tioned Substance		
2					
3 4	Chemical Names: Hypochlorous acid, hypochloric(I) acid, chloranol,	14 15	Clean Smart, FloraFresh® Floral Quality Care Solution, HSP2O Pro Disinfectant		
5	hydroxidochlorine		CAS Numbers: 7790-92-3		
6 7	Other Name: Hydrogen hypochlorite, Chlorine hydroxide		Other Codes: European Community Number- 22757, IUPAC-Hypochlorous acid		
8	Trade Names: Oculus Puracyn Antimicrobial		List other codes: PubChem CID 24341		
9 10	Skin and Wound Cleanser, Vashe® Wound Therapy Solution (OTC use), Vashe® Wound		InChI Key: QWPPOHNGKGFGJK- UHFFFAOYSA-N		
11 12 13	Therapy Solution (Professional use), NixallTMWound and Skin Care (OTC and Professional use), Excelyte VET,		UNII: 712K4CDC10		
16	Summary	of Pet	titioned Use		
17 18 19 20 21 22 22 23 224 25 26 27 28 29 33 33 34 33 35 33 36	for wound care, post milking teat sanitation, dern Recently, the National Organic Standards Board (electrolyzed water for use in organic crop and live separate recommendations for its addition to the 205.605(b). In a memorandum responding to this forward with a proposed rule for public commenterport, a proposed rule has not yet been published materials including HOCl respectively in crops for disinfecting and sanitizing facilities and equipmented contact surfaces where residual chlorine remains 4 milligrams/liter, the previous NOSB recommented in a previous technical evaluation report requestions and previous technical evaluation report requestions.	olyzed vestock or organization of to add to me and below dation uested rt and	water to section 205.603 of the National List: a production. HOCl is petitioned for use as a anic livestock production. Direct HOCl contact in commercial veterinary and farm products used gical disease and the treatment of eye irritation. Considered a petition to allow HOCl from production and handling and passed three hal List in sections 205.601(a)(2), 205.603(a)(7) and mendation, NOP indicated its intent to move d HOCl to the National List. At the time of this le these sections apply to the use of chlorine arvest use and sprout production, in livestock for in handling for disinfecting and sanitizing food the maximum residual disinfectant limit, currently as did not consider the direct contact of HOCl with		
38	Characterization of Petitioned Substance				
39	Composition of the Substance:				
40 41 42 43 44 45	HOCl is an oxyacid of chlorine containing monovalent chlorine that acts as an oxidizing or reducing agent (NCBI, 2017). A 2015 technical report on HOCl (handling/processing uses) is available at the NOP Petitioned Substances Database. HOCl is a chlorine releasing agent. It is one of three forms of aqueous chlorine: chlorine gas (Cl ₂), HOCl and hypochlorite ion (OCl-). Mostly HOCl is present in aqueous solutions pH between 2 - 7.5 at 25°C (Kettle et al., 2014).				
46					

Source or Origin of the Substance:

47

50

52

53

58

61

64

67 68

69 70

74

48 HOCl is produced by electrolyzing a brine solution made with purified water and sodium chloride. In

49 general, the brine is submitted to direct electrical current as it flows through a cell that allows the physical

separation of solutions around the positive and negative electrodes. Separation is usually achieved with a

semi-permeable membrane. At pH 3.5 - 6.5 (30°C), greater than ninety percent of the chlorine species

present at the anode is HOCl. If the flow cell is constructed properly HOCl can be removed from the anodic

compartment as it is produced. Sodium hydroxide produced at the cathode is separately removed.

Dakin's solution, a solution containing a low concentration of sodium hypochlorite (household bleach),

boric acid and sodium bicarbonate was adopted in the early 1900s and is still used as an effective

56 microbiocide and disinfectant for burns and wounds (Dakin, 1915a,b). HOCl at a concentration of 0.3

57 percent is the predominant chlorine releasing agent present in Dakin's solution. Dakin's solution is not

produced electrolytically. It is effective in killing *Staphylococcus* spp., *Streptococcus* spp. and *E. coli* (Smith et

59 al., 1915). The effectiveness of Dakin's solution and the presence of HOCl is pH dependent. Although

Dakin's solution is not produced from electrolyzed water, it is still a source of pure HOCl.

Properties of the Substance:

There is a pH and temperature dependent equilibrium between the three aqueous chlorine species, Cl₂,

OCI and HOCI. This relationship for selected values of pH and temperature is shown for HOCI and OCI-

(Table 1). Of the three aqueous species, HOCl is the primary bactericidal agent (White, 1972). As the pH

65 increases the percentage of HOCl decreases with an increase in the percentage of OCl-. With increasing

ionic strength, the percentage of HOCl present in solution also decreases. At pH 7.5, with other ions such

as phosphate present in solution the amount of HOCl in solution can be as low as 51%, balanced by 49%

OCl⁻ (White, 2011).

Table 1 Percentage of chlorine present as HOCl as a function of pH and temperature (assuming zero ionic strength and considering HOCl and OCl as the only chlorine species present)

рН	Percent HOCl							
	0°C	5°C	10°C	15°C	20°C	25°C	30°C	
5.0	99.85	99.82	99.80	99.77	99.74	99.71	99.68	
5.5	99.53	99.45	99.36	99.27	99.18	99.09	99.00	
6.0	98.53	98.27	98.00	97.72	97.45	97.18	96.92	
6.5	95.48	94.72	93.94	93.14	92.35	91.58	90.86	
7.0	86.99	85.02	83.05	81.11	79.24	77.48	75.86	
7.5	67.89	64.23	60.78	57.59	54.69	52.11	49.84	
8.0	40.06	36.21	32.89	30.04	27.63	25.60	23.91	
8.5	17.45	15.22	13.42	11.95	10.77	9.81	9.04	
Adapte	Adapted from Black and Veatch Corporation, 2011							

Specific Uses of the Substance:

71 HOCl is the active microbiocidal ingredient in a number of veterinary preparations used for treatment of

72 infectious keratoconjunctivitis (pinkeye), burns, wounds and mastitides. Keratoconjunctivitis is an

73 inflammation of the covering membrane of the eye, including the orbit and the inner surface of the eyelids.

This inflammation typically extends below the conjunctival layer (Farley, 1941). There are several causative

July 12, 2017 Page 2 of 12

- 75 agents. In cattle the most significant cause is the bacterium Moraxella bovis, and to a lesser extent Chlamydia
- 76 spp., Neisseria catarrhalis, and Mycoplasm spp. In sheep, the causes are Rickettsia conjunctivae, Neisseria
- 77 catarrhalis, Mycoplasma conjunctivae, Acholeplasma oculi and Chlamydia spp. In goats, the cause is Rickettsia
- 78 conjunctivae. In pigs, the cause is Rickettsia spp. In horses, the cause is frequently viral, but bacteria such as
- 79 *Moraxella spp.* have been found associated with the inflammation (Radostits et al., 1994). Treatments that
- include HOCl are solutions, washes, teat dips and gels. They may be buffered or osmotically balanced.
- HOCl is used at a relatively low concentration in these treatments, 0.01-0.02%.

Approved Legal Uses of the Substance:

- 83 Veterinary HOCl solutions are regulated by the US Food and Drug Administration (FDA) through the
- premarket notification or 510 (k) programs. The procedure for 510 (k) submission is listed at
- 85 §21 CFR 807.21. The 510(k) is a manufacturer's self-certification of the safety and efficacy of the product
- 86 that otherwise does not require licensing. The FDA reviews and evaluates the product and determines 1)
- 87 that a premarket approval is necessary or 2) that a premarket approval is not required. After the
- 88 determination is made a letter is sent to the applicant allowing the product to be marketed. Solutions of
- 89 hypochlorous acid for use in treatment of eye irritation and wound care are already legally marketed. New
- 90 products such as those mentioned in the petition are considered substantially equivalent by the FDA and
- 91 do not require premarket approval.
- 92 A petition has been received by the USDA to add veterinary HOCl solutions for organic livestock
- 93 production to the National List.

Action of the Substance:

- 95 Neutrophils are white blood cells involved in the mammalian antibacterial immune response. They contain
- an enzyme called myeloperoxidase that reacts with hydrogen peroxide also produced by the neutrophil at
- 97 sites of bacterial infection. Myeloperoxidase catalyzes the formation of hypohalous acids from hydrogen
- 98 peroxide (Mika and C, Guruvayoorappan, 2011). HOCl is the major product. It is antimicrobial in vitro,
- 99 although the actual antimicrobial process requires the conversion of HOCl to chloramine T which serves as
- the antimicrobial reactive oxidative species. Chloramine T forms when HOCl reacts with amine groups
- 101 (Amulic et al., 2012).
- 102 HOCl rapidly kills the causative agents of infectious keratoconjunctivitis, wound infections and mastitides
- 103 (Gard et al. 2012; Amulic et al., 2012). In addition to its microbiocidal effect, HOCl can have an anti-
- inflammatory effect by 1) reducing histamine activity, 2) reducing leukotriene activity, 3) increasing TGF-
- beta activity, 4) increasing growth factor synthesis and 5) decreasing metalloprotease 7, collagenase and
- gelatinase activity (Pelgrift and Friedman, 2013).

Combinations of the Substance:

- 108 Commercially available treatments contain products from the production of electrolyzed water, including
- sodium chloride (NaCl), HOCl and sodium hypochlorite, but may also contain added pH buffers such as
- sodium phosphate (NaH₂PO₄/ Na₂HPO₄), sodium sulfate and sodium bicarbonate, and colloidal gels such
- as lithium magnesium sodium silicate depending upon the tissue to be treated. A teat dip product that
- releases HOCl consists of two dissolved 2.5 g sodium dichloroisocyanuarate tablets dissolved in one liter of
- water with an available chlorine concentration of 2800 ppm at pH 5.5-6.0 (Bodie et al., 1995).

Limited Scope Questions for Hypochlorous Acid to be Used Livestock Production

For this limited scope report, the NOSB Livestock Subcommittee has requested that the report respond to the specific questions listed below.

117

114

107

82

94

- Evaluation Question #1: What is the efficacy of hypochlorous acid for pinkeye and wound treatment, relative to other products (both synthetic and on the National List)?
- relative to other products (both synthetic and on the National List):
- 120 HOCl is available in several commercial presentations for veterinary treatment of pink eye and wounds. A
- 121 number of studies with these preparations have shown the efficaciousness of HOCl for pinkeye, wounds,
- burns and mastitides (Gard et al., 2016; Pegrift and Friedman, 2013; Hua et al., 2003). In addition to HOCl
- and hypochlorite, these products can contain sodium phosphate or silicate in the case of colloidal burn

July 12, 2017 Page 3 of 12

- 124 treatments. HOCl and hypochlorite salts are not currently on the National List for treatment of pink eye
- 125 and wounds. Sodium phosphates and aqueous potassium silicate are approved for use respectively in
- 126 organic handling (§205.605(b) and crop production (§205.601(e)(2)). Sodium phosphate or sodium silicate
- 127 may be allowed as excipients if their use complies with the requirements at section 205.603(f).
- 128 HOCl is a chlorine releasing agent (CRA). It is considered the active moiety responsible for bacterial
- 129 inactivation by CRAs. CRA microbiocidal activity is greatest when the percentage of undissociated HOCl is
- 130 highest. At a concentration of 50 micromoles (μ M)/liter (I) (2.6 ppm), HOCl produces deleterious effects on
- 131 bacterial DNA resulting in the formation of chlorinated derivatives of nucleotide bases and complete
- 132 inhibition of Escherichia coli growth. Concentrations greater than 5 millimoles/1 (260 ppm) have been found
- 133 to disrupt oxidative phosphorylation and other membrane-associated activity (McDonnell and Russell,
- 134
- 135 The primary cause of infectious bovine keratoconjunctivitis (pinkeye) is the bacteria Moraxella bovis. It is a
- gram negative, aerobic, oxidase positive diplococcus (Holt, 1994). The bacterial pili and the enzyme 136
- hemolysin have both been established in disease pathogenesis. The pili help to attach bacteria to the 137
- 138 cornea, while hemolysin is linked to cell membrane disruption. In addition, the host immune response
- 139 against the bacteria can be prolonged due to an immunoevasive process called phase variation. Phase
- 140 variation permits the bacterial to rearrange its genes for pilus formation producing a changing immune
- 141 signature for the largest bacterial organelle. Without a solid short term, immune response from the animal
- 142 pinkeye can be exacerbated and prolonged (Marrs et al., 1988). Vaccines used for pink eye must contain all
- 143 of the variant pilus types to be effective, thus many bacterins or killed vaccines commonly used in organic
- 144 production are not always effective (McConnell et al., 2008; O'Conner et al., 2011). However, without other
- 145 treatment options, many organic producers still choose to vaccinate. Vaccines are on the National List at
- section 205.603(a)(4) as "Biologics Vaccines." 146
- 147 A commercial veterinary spray treatment containing 0.009% HOCl, sodium chloride and phosphate buffer
- 148 was examined by a group at Auburn University. The study consisted of thirty calves randomized into three
- 149 groups. In groups one and two a corneal lesion was also induced into the left eye and the same eye was
- 150 then infected with *Moraxella bovis* to induce pinkeye. The right eye of these two groups served as a control.
- 151 No corneal lesion was induced. Nor was it infected with *M. bovis*. A third group did not receive a corneal
- 152 lesion in either eye, but was inoculated with M. bovis in the left eye. Beginning 24 hours after inoculation
- 153 with M. bovis, calves in group 1 were treated with 0.009% HOCl for a total of 2 milliliters per application (3
- 154 sprays) twice daily for ten days. Calves in group 2 were treated 24 hour post M. bovis inoculation with 0.9%
- 155 sterile saline solution for a total of 2 milliliters per application (3 sprays) twice daily for ten days. Following
- 156 inoculation with M. bovis, calves in group three were not treated. Both eyes in all three groups were
- 157 evaluated for evidence of pain and ocular discharge associated with the clinical signs of infectious bovine
- keratoconjunctivitis (IBK) throughout the study. In addition, blood and liver biopsy samples were collected 158
- 159 to determine if chlorine levels increased internally. All calves in groups one and two developed corneal
- ulcerations and clinical signs consistent with IBK in the left eye. No clinical signs were observed at any 160
- 161 time during the study in group three or the right eye of groups one and two. After ten days, all of the
- 162 animals in groups one and three were culture negative for M. bovis. Average healing time of corneal lesions 163 was 3.7±1.2 days for group one and 8.3±3.7 for group 2 (p<0.0002). In this study, topical ophthalmic
- 164 application of HOCl was effective in treating experimentally induced pinkeye. No additional chlorine
- 165 accumulated internally in any of the test animals. Pain experienced by the calves as a result of the pinkeye
- infection was also much less in the group treated with HOCl (Gard et al., 2016). 166
- There are a number of studies in both human and rat showing positive results for healing and pain relief 167
- 168 from the use of HOCl solutions for debridement and maintenance of burns and wounds (Selkon et al.,
- 169 2006; Liden, 2013; Robson, 2007; Nakae and Inaba, 2000; Hua et al., 2003; Sakarya et al., 2014). In the case of
- 170 colorectal surgery, however; in which no significant difference was observed between using HOCl and
- saline, HOCl is not recommended (Takesue et al., 2011). When used for veterinary wound treatment, 171
- 172 solutions containing a low concentration (0.11-0.12 %) of HOCl produced either by electrolysis of water or
- 173 dilution of sodium hypochlorite (Dakin's solution) provided a reduction in bacterial infection and
- 174 improved wound healing (Krahwinkel and Boothe, 2006; Ramey and Kinde, 2015).
- 175 During wound healing, there is a pH shift from acidic, which is the pH of normal skin to slightly alkaline,
- 176 resulting from the myriad of biochemical reactions facilitating degradation of dead tissue, the intercalation

July 12, 2017 Page 4 of 12

- of the wound with new structures of extracellular matrix and the reassembly of connective tissue and
- epithelium (Schnieder et al., 2007). The intervention of phagocytic white blood cells (neutrophils) during
- this process includes the release of a chlorine to kill bacteria and fungi. An acidic pH adjustment also
- occurs during this particular step which favors myeloperoxidase mediated production of the chlorine
- releasing agent, HOCl (Pullar et al., 2000). Cellular antioxidants are produced to control excessive HOCl
- 182 production and prevent tissue damage (Mika and Guruvayoorappan C, 2011).

183 <u>Evaluation Question #2:</u> What items are being used for pink eye and wound treatment on organic

184 **farms?**

- Face flies are known to carry and transmit the bacteria *Moraxella bovis*, the etiological agent for infectious
- bovine keratoconjunctivitis (Berebile and Webber, 1981; OConnor et al., 2012). Controlling flies helps to
- 187 reduce the risks of disease spread between animals in a herd. The face fly, *Musca autumnalis* (Diptera:
- 188 Muscidae), feeds at the eyes and faces of cattle and horses in the temperate regions of the northern
- 189 hemisphere. Feeding flies probe with their proboscis and consume secretions or discharges from their
- 190 hosts' eyes, nostrils, mouths, vulvae, teats, and other body parts. Sharp spines surrounding the fly's mouth
- may cause superficial lesions increasing the likelihood of infection. Eggs and larvae occur exclusively in the
- dung pats of cattle and bison (Krafsur and Moon, 1997). Pinkeye was enzootic in North America before the
- face fly was introduced, but its prevalence has increased since the fly arrived. Face fly abundance correlates
- 194 positively with disease incidence (Hall, 1984). Face flies are attracted to a number of substances including
- cattle fed a diet of alfalfa (Pickens and Miller, 1980). Vespid wasps such as Vespula germanica (Fabricius)
- have been identified as a predator of face flies. These and other predatory wasps can also be used to control
- 197 maggot fly larvae in dung (Schidtmann, 1977; Skovgard, 2004). Ground phosphate rock can be scattered
- over gutter manure and manure heaps to destroy fly larva (Surface, 1915). Fly traps can be used for
- removal of flies from the animals or their housing (Denning et al, 2014; Kaufman et al., 2005). Other organic
- 200 production methods for fly control can include the use of cow masks, cleaning out box stall buildup
- 201 (especially the near the corners), having chickens peck away the manure paddies after animals have been
- 202 through a paddock and/or having hogs root through the paddies. Frequent applications of extracts, and
- essential oils as well as sulfur dusting may also provide some relief (An M.R.C.V.S., 1915).
- 204 Flies are generally more numerous in tall grass. Trimming tall grass, and foxtails can serve to reduce fly
- 205 populations (Bersford and Sutcliffe, 2008). Trimming tall grass also help to prevent scratches to the eyes of
- 206 feeding animals. Reducing moisture in the animal's environment helps to reduce fly numbers. Sprinkling
- field lime or diatomaceous earth on the animals helps to keep them dry. Because pinkeye is infectious
- affected animals should be separated if possible. However, in some cases, M. bovis may infect animals in a
- 209 carrier state where no clinical signs appear, but the animal remains infectious and bacterial transmission is
- 210 possible. Here diagnosis is important.
- 211 Topical treatment is recommended for both pinkeye and wounds. Repeated and frequent applications of
- various combinations of isotonic saline solution, calendula, garlic, echinacea, cochlearia, honey (wounds),
- sugar (wounds), essential oils, eyebright, goldenseal, colloidal silver, breast milk cod liver oil, Aloe vera and
- coconut oil have been used as washes and topical treatments (Zinke, 2010; Menendez et al., 2007; Swaim
- and Lee, 1987). Many of these substances are microbiocidal and serve to kill or control the bacterial
- infection (Calvo and Czvero, 2016). For pinkeye, anything that soothes the eye is also helpful. Dairy cattle
- and calves are generally easier to treat, since restraint may be required for application. No one method is
- 218 preferable and efficacy is not assured. In the case of wounds, and in addition to washing the wound,
- disinfection is generally indicated (Krahwinkel and Boothe, 2006).
- 220 Kelp meal is often fed to organic dairy cattle serving as a mineral and vitamin source (Hardie et al., 2014)).
- 221 In addition, vitamins A, D, and E are helpful in maintaining the livestock immune system and eye health
- 222 (Anonymous, 1976). Adequate levels of trace minerals such as copper and selenium are also indicated for a
- properly functioning immune system. Because sunlight and ultraviolet light aggravates pinkeye, animals
- should be sequestered from sunlight and ultraviolet. It is best to let them out at night for grazing if this is
- possible.
- 226 As of April 5, 2017, seven licensed *M. bovis* bacterin vaccines, one conditionally licensed *M. bovoculi*
- bacterin vaccine, three combination bacterin-toxoid vaccines containing M. bovis, one M. bovis bacterin for
- 228 further manufacture and one M. bovis killed culture for further manufacture are listed in the USDA Animal
- 229 Plant Health Inspection Service's <u>USDA Veterinary Biological Products</u>. These are also listed in Table 2.

July 12, 2017 Page 5 of 12

Bacterins are normally acceptable for use in Organic Livestock Production. Bacterins are simply bacteria killed with formaldehyde. The formaldehyde is dried off, and an adjuvant is added to stimulate the immune response. Bacteria used for the bacterin may be a naturally occurring isolate or it may be genetically modified. Because M. bovis exhibits phase variation, several isolates are combined into the vaccine to cover possible immune variation. Furthermore, M. bovoculi was recently recognized as an additional causal bacterium for bovine pinkeye. A vaccine for this M. bovoculi is now also available (OConnor et al., 2012). Vaccines are a useful tool for reducing production losses (McConnel et al., 2008). However, the highly variable immunogenic profile of *M. bovis* has complicated the use of vaccines. Many producers use them, but they do not always work. Even a vaccine produced from isolates from the same farm where it was used was not very efficacious, raising a concern that even autogenous vaccines are often ineffective in controlling naturally occurring IBK (OConnor et al., 2011). Vaccinations should be administered well in advance (ideally at least four weeks) of the anticipated summer onset of pinkeye, so that cattle will have enough time to mount an effective immune response following vaccination. Young animals tend to be most affected, and should be a part of the vaccination program.

243244

230

231

232233

234

235

236

237238

239

240

241242

Table 2 Vaccines for Infectious Bovine Keratoconjunctivitis					
Type of Vaccine	USDA Vaccine Code	Producer	Description		
Moraxell bovis bacterin	2772.00	1, 2, 3, 4	Formaldehyde inactivated, multiisolate		
	2772.03	5	bacterin. Contains an adjuvant to stimulate the immune response. Different isolates,		
	2772.04	5	numbers of isolate, adjuvants, etc.		
	2772.10	6			
Moraxella bovoculi bacterin	2A77.00	4	For pinkeye caused by M. boviculi		
Clostridium Chauvoei-	7425.01	1	Multi valent bacterin-toxoid vaccine,		
Septicum-Novyi-Sordellii- Perfringens Types C & D-	7425.02	5	contains IVI. DOVIS Dacterin.		
Moraxella Bovis Bacterin- Toxoid	7425.03	5			
Moraxell bovis bacterin	B772.00	4	for further manufacture		
Moraxell bovis killed culture	B772.01	4			

¹Boehringer Ingelheim Vetmedica, Inc., Divisions: Bio-Ceutic, Anchor, 2621 North Belt Highway, St. Joseph, MO 64506 (124, 124A)

July 12, 2017 Page 6 of 12

² Elanco US Inc. 196, Subsidiaries: Lohmann Animal Health International, Elanco US Inc., 1447 140th Street, Larchwood, IA 51241, 196

³ Novartis Animal Health US, Inc., 1447 140th Street, Larchwood, IA 51241-9778 303

⁴ Addison Biological Laboratory, Inc., Route 3 Box 90-B, Fayette, MO 65248 355

⁵ Intervet Inc. 165A, Divisions: Merck Animal Health, Merck Sharpe and Dohme (MSD), 21401 West Center Road, Elkhorn, NE 68022

⁶ SolidTech Animal Health, Inc. 604, 812 NE 24th Street, P.O. Box 790, Newcastle, OK 73065-0790

Hair color, age, host behavior such as comparisons of movement, grazing and lying down and aggregation of host animals have been reported to influence cattle-host selection by face flies and predilection for pinkeye. Breeds which lack pigment on their eyelids (Herefords, Hereford crosses, Charolais and some Holsteins) are hypothesized to be more susceptible to pinkeye than dark-hided cattle. Although lighter hair and eye color suggests more susceptibility, hair color and eye pigment are likely not to be the only selective criteria. The breed that appears to be most susceptible to face flies and pinkeye is the Hereford. While

tropically adapted breeds such as Brahmin are less susceptible. (Steelman et al., 1993, Snowder et al., 2005). Genetic tracking of breed susceptibility to pinkeye has followed the quantitative trait locus (QTL). A QTL is a section of genetic information (DNA) on a chromosome (the locus) in the genome that correlates with variation in a phenotype (the quantitative trait). Usually the QTL is nearby on the chromosome, or contains, the genes that control the specific phenotype. QTLs are mapped by identifying the molecular biomarkers correlated with an observed trait. This is sometimes an early step in identifying and sequencing the actual genes that cause the trait variation (Miles and Wayne, 2008). QTLs have been identified for infectious bovine keratoconjunctivitis (IBK) on chromosome 1 and 20 associated with the probability of contracting this disease (Casa and Stone, 2006). The QTL on chromosome 1 may have a link to the bovine macrophage activity, while the QTL on chromosome 20 may be associated with the cellular or non-specific immune response (Garcia et al., 2010; Casa and Stone, 2008).

Wound care begins with direct pressure to the wounded site. This helps to stop the bleeding. Ice also constricts vessels and will slow bleeding. Irrigation with saline, dilute disinfectant or water is next to wash away contamination including bacteria and stimulate healing. Both iodine and chlorhexidine are allowed for use as disinfectants when other disinfection methods are not expected to work. Iodine is allowed for use as a disinfectant and a topical treatment. Aqueous iodine is antibacterial, but at higher concentration is deleterious to tissues in vivo and may potentiate infection. Iodine is more effective at lower concentrations, 0.1% or 1%, rather than the 10% solution that is normally supplied (Swaim and Lee, 1987). Chlorhexidine is allowed for surgical procedures conducted by a licensed veterinarian. Chlorhexidine is an effective antimicrobial. Wounds irrigated with 0.1%, 0.25% or 1% chlorhexidine did better than wounds respectively irrigated with 0.1%, 0.25% and 0.5% iodine and polyvinylpyrrolidone. However, a correlation between chlorhexidine use and joint inflammation has been noted. Although not approved for organic use, Dakin's solution, a dilute sodium hypochlorite solution (0.005%) has been found to be more effective at killing Staphyloccus aureus than chlorhexidine (Swain and Lee, 1987). Externally copper sulfate is antiseptic, astringent, caustic germicidal, fungicidal and viricidal. It can be used in solution for wound debridement and applied as a powder to wounds and as an antiseptic post surgical treatment (Mouli, 2005). Ringer's solution is effective for wound debridement and some relief from pain (Colegrave et al., 2016). Ringer's solution typically contains sodium chloride, potassium chloride, calcium chloride and sodium bicarbonate, with the last used to balance the pH.

Evaluation Question #3: What natural items could be used?

Natural items are products that come from plants, animals, or minerals. There are many remedies that have not been experimentally tested. Several organically produced combinations are available commercially. Table 3 provides a non-exhaustive list of known natural products that may be used in pinkeye and/or wound treatment. Many of these treatments may be used under veterinary supervision. While others can

be practically applied without a veterinarian's help.

252

253254

255

256257

258

259260

261

262

263264

265266

267

268

269

270

271272

273

274

275

276

277

278

279

280281

287 288

289

290

291

292

293

294

295

296

<u>Evaluation Question #4:</u> What does the National List currently allow for pink eye and wound treatment?

One of the most compelling aspects of organic farming is system health. Organic health management includes the creation of a complete system for sustainable livestock welfare minimizing vectors of disease and ensuring livestock are provided proper nutrition and environment to build a strong natural immune repertoire when stress and pathogens occur (Coffey and Baier, 2012). Water washing, and isotonic saline washing are allowed for organic livestock production. Non-synthetic (natural) herbal microbiocide decoctions are not specifically listed in the National List, but are allowed in Organic Livestock production. Ethanol and isopropanol are permitted as topical disinfectants (7 CFR 205.603(a)(1)(i)) and can be used for wound care. Chlorhexidine is also indicated and allowed for surgical procedures conducted by a veterinarian (7 CFR 205.603(a)(6)). Biologics—Vaccines (7 CFR 205.603(a)(4)) for pinkeye are allowed for

veterinarian (7 CFR 205.603(a)(6)). Biologics – Vaccines (7 CFR 205.603(a)(4)) for pinkeye are allowed forganic livestock production. For wound care, both hydrogen peroxide hydrogen peroxide (7 CFR

July 12, 2017 Page 7 of 12

Table 3 Natural substances used for pinkeye and wound treatment							
<u>Plant or substance</u>	Classification Nomenclature	Application					
Seabuckthorn	Hippophae rhamnoides	Wounds ³					
Parasitic wasps	Spolangia cameroni	Pinkeye ³					
Kiwi fruit	Actinidia deliciosa	Wounds 8					
Aloe	Aloe vera	Pinkeye, wounds 14,5					
Calendula	Calendula officinalis	Pinkeye 1					
Chamomile	Chamoemelum nobile	Pinkeye 1					
Chicory	Cichorium intybus	Pinkeye 1					
St. John's Wort	Hypericum perforatum	Pinkeye 1					
Olive	Olea europaea	Pinkeye 1					
White poplar	Populus alba L.	Pinkeye 1					
Rose	Rosa agrestis	Pinkeye 1					
Elder	Sambucus nigra L. ssp. nigra	Pinkeye 1					
Navelwort	Umbilicus rupestris	Pinkeye 1					
Mullein	Verbascum sinuatum L.	Pinkeye 1					
Veronica	Veronica spp.	Pinkeye 1					
Physic nut	Jatropha curcas	Wounds 6					
Bacterial predators	Bdellovibrio bacteriovorus	Pinkeye ²					
Brown kelp alginates	Ascophyllum nodosum	Wounds ⁵					
Honey		Wounds 5					
Sugar		Wounds 5					
Pineapple Fruit Enzymes (bromelain)	Ananas comosus	Wounds 7					
Omentum		Wounds 5					
Chitosan		Wounds 5					
Platelet Gel		Wounds 5					
Pink Trumpet Tree	Tabebuia avellanedae	Wounds 9					
Brazialian Pepper Tree	Schinus terebinthifolius	Wounds 9					
Siam Weed	Chromolaena odorata	Wounds 10					
1							

 $^1\text{Calvo}$ and Cavero, 2016; $^2\text{Boileau}$ et al., 2011; $^3\text{Skovgard}$, 2004; $^4\text{Swaim}$ and Lee, 1987; $^5\text{Krahwinkel}$, D.J. and Boothe; $^6\text{Thomas}$ et al., 2008; $^7\text{Rosenberg}$ et al., 2004; $^8\text{Hafzei}$ et al., 2010; $^9\text{Lipinski}$ et al., 2012; $^{10}\text{Vijayaraghavan}$ et al., 2017.

July 12, 2017 Page 8 of 12

- 299 205.603(a)(11)) and iodine (7 CFR 205.603(a)(14)) are allowed. In addition, vitamins (7 CFR 205.603(d)(2))
- and mineral supplements (7 CFR 205.603(d)(3)), e.g. vitamin A, D, and E, may be added the diet to improve
- 301 the immune response and the immune health of the eye. The producer of an organic livestock operation
- must not withhold medical treatment from a sick animal in an effort to preserve its organic status. All
- 303 appropriate medications must be used to restore an animal to health when methods acceptable to organic
- production fail. Livestock treated with a prohibited substance must be clearly identified and shall not be
- sold, labeled, or represented as organically produced (7 CFR 205.238(c)(7)).

306 References

- Amulic, B., Cazalet, C., Hayes, G.L., Metzler, K.D. and Zychlinsky, A. (2012) Neutrophil function: from
- mechanisms to disease, Annu. Rev. Immunol., 30, pp. 459–489.
- 309 An M.R.C.V.S. (1915) The Farm Vet, McDonald and Martin, London.
- 310 Anonymous (1976) More minerals and vitamin A, less face flies: check pinkeye, Livestock Breeders Journal,
- 311 19:7, pp. 177-179.
- 312 Beresford, D.V. and Sutcliffe, J.F. (2008) Stable fly (Stomoxys calcitrans: Diptera, Muscidae) trap response to
- changes in effective trap height caused by growing vegetation, Journal of Vector Ecology, 33:1, pp. 40-45.
- Berkebile, D.R, Hall, R.D. and Webber, J.J. (1981) Field association of female face flies with Moraxella bovis,
- an etiological agent of bovine pinkeye, J. Economic Entomology, 74:4, pp. 475-477.
- 316 Black and Veatch Corporation (2011) White's Handbook of Chlorination and Alternative Disinfectants,
- 317 John Wiley and Sons.
- Bodie, R.L., Nickerson, S.C., Doyle, M.G., and McGuire, H.J. (1995) Efficacy of a new hypochlorous acid-
- 319 releasing teat dip against Staphylococcus aureus and Streptococcus agalactiae under conditions of experimental
- challenge, National Mastitis Council Meeting Proceedings, 34, pp. 156-157.
- 321 Boileau, M.J., Clinenbeard, K.D. and Iandolo, J.J. (2011) Assessment of *Bdellovibrio bacteriovirus* 109J killing
- 322 of Moraxella bovis in an in vitro model of infectious bovine keratoconjunctivitis, The Canadian Journal of
- 323 Veterinary Research, 75, pp. 285-291.
- 324 Calvo, M.I. and Cavero, R.Y. (2016) Medicinal plants used for ophthalmological problems in Navarra
- 325 (Spain), Journal of Ethnopharmacology, 190, pp. 212–218.
- 326 Calvo, M.I. and Czvero, R.Y. (2016) Medicinal plants used for ophthalmological problems in Navarra
- 327 (Spain), Journal of Ethnopharmacology, 190, pp. 212–218.
- Casas, E. and Stone, R. T. (2006) Putative quantitative trait loci associated with the probability of
- 329 contracting infectious bovine keratoconjunctivitis, Journal of Animal Science, 84:12, pp. 3180-3184.
- Casas, E. and Stone, R. T. (2008) A putative quantitative trait locus on chromosome 20 associated with
- bovine pathogenic disease incidence, Journal of Animal Science, 86, pp. 2455-2460.
- 332 Coffey, L. and Baier, A.H. (2012) Guide for Organic Livestock Producers, ATTRA, National Center for
- 333 Appropriate Technology, USDA, NOP.
- Colgrave, M., Rippon, M.G. and Richardson, C. (2016) The effect of Ringer's solution within a dressing to
- elicit pain relief, Journal of Wound Care, 25:4, pp. 184-190.
- Dakin, H.D. (1915a) On the use of certain antiseptic substances in the treatment of infected wounds, British
- 337 Medical Journal, 2:2852, pp. 318-320.
- Dakin, H.D. (1915b) The antiseptic action of hypochlorites: the ancient history of the "new antiseptic,"
- 339 British Medical Journal, 2:2866, pp. 809-810.
- Denning, S.S., Washburn, S.P. and Watson, D.W. (2014) Development of a novel walk-through fly trap for
- the control of horn flies and other pests on pastured dairy cows, J. Dairy Sci., 97, pp. 4624–4631.
- Farley, H. (1941) Keratitis: the virulence, transmissibility and coarse of bovine 'pinkeye,' Journal of the
- 343 American Veterinary Association, 3, pp. 74-76

July 12, 2017 Page 9 of 12

- Garcia, M.D., Matukumalli, L., Wheeler, T.L., Shackelford, S.D, Smith, T.P.L., and E. Casas, E. (2010)
- Markers on bovine chromosome 20 associated with carcass quality and composition traits and incidence of
- contracting infectious bovine keratoconjunctivitis, Animal Biotechnology, 21, pp. 188–202.
- Gard, J., Taylor, D., Maloney, R., Schnuelle, M., Duran, S., Moore, P., Justus, W., Walz, P., Stockle, R.,
- Rodning, S., DeGraves, F., van Santen, E., Edmonson, M. and O'Conner, A.M. (2016) Preliminary
- evaluation of hypochlorous spray for treatment of experimentally induced infectious bovine
- keratoconjunctivitis, The Bovine Practioner, 50:2, pp. 180-189.
- Hafezi, F., Rad, H.E., Naghibzadeh, B., Nouhi, A. and Naghibzadeh (2010) Actinidia deliciosa (kiwifruit), a
- new drug for enzymatic debridement of acute burn wounds, Burns, 36, pp. 352–355.
- 353 Hall, R.D. (1984) Relationship of the face fly (Diptera: Muscidae) to pinkeye in cattle: a review and
- synthesis of the relevant literature, J. Med. Entomol., 21, pp. 361–365.
- Hardie, C.A., Wattiaux, M., Dutreuil, M. Gildersleeve, R. Keuler, N.S. and Cabrera, V.E. (2014) Feeding
- 356 strategies on certified organic dairy farms in Wisconsin and their effect on milk production and income
- 357 over feed costs, J. Dairy Sci., 97, pp. 4612-4623.
- 358 Holt, J.G. (1994) Bergey's Manual of Determinative Bacteriology, LWW.
- 359 Hua, X., Zheng, Y-I., Nakae, H. and Han, Z-G (2003) Effect of electrolyzed oxidizing water and
- 360 hydrocolloid dressings on excised burn-wounds in rats, Chinese J. Traumatology, 6:4, pp. 234-237.
- 361 Hua, X., Zheng, Y-j., Nakae, H. and Han, Z-G. (2003) Effects of electrolyzed oxidizing water and
- 362 hydrocolloid occlusive dressings on excised burn-wounds in rats, Chinese Journal of Traumatology, 6:4,
- 363 pp.234-237.
- 364 Kaufman, P.E., Rutz, D.A. and Frisch, S. (2005) Large Sticky Traps for Capturing House Flies and Stable
- Flies in Dairy Calf Greenhouse Facilities, J. Dairy Sci., 88, pp. 176–181.
- Kettle, A.J., Albrett, A.M., Chapman, A.L., Dickerhof, N., Forbes, L.V., Khalilova, I. and Turner, R. (2014)
- Measuring chlorine bleach in biology and medicine, Biochimica et Biophysica Acta, 1840, pp. 781–793.
- 368 Kidwell, B. (2016) Put an end to pinkeye, Progressive Farmer, March.
- Krafsur, E.S. and Moon, R.D. (1997) Bionomics of the face fly, Musca autumnalis, Annu. Rev. Entomol., 42,
- 370 pp. 503-523.
- 371 Krahwinkel, D.J. and Boothe, H.W. (2006) Topical and systemic medications for wounds, Vet. Clinics. Small
- 372 Anim., 36:4, pp. 739-757.
- 373 Krahwinkel, D.J. and Boothe, H.W. (2006) Topical and systemic medications for wounds, Vet Clinics Small
- 374 Anim., 36, pp. 739-757.
- Liden, B.A. (2013) Hypochlorous Acid: Its Multiple Uses for Wound Care, Ostomy Wound Management,
- 376 September, pp. 8-10.
- 377 Lipinski, L.C., Wouk, P.D.F., de Silva, N.L., Perotto, D. and Ollhoff, R.D. (2012) Effects of 3 Topical Plant
- Extracts on Wound Healing in Beef Cattle, Afr J Tradit Complement Altern Med., 9:4, pp. 542-547.
- Marrs, C.F., Ruehl, W.W., Schoolnik, G.K. and Falkow, S. (1988) Pilin gene phase variation of Moraxella
- 380 bovis is cause by an inversion of the pilin genes, Journal of Bacteriology, 170:7, pp. 3032-3039.
- McConnel, C.S., Shum, L., Gleeson, B.L. and House, J.K. (2008) Serologic cross-reactivity of Australian
- 382 McConnell, C.S., Shum, L., Gleeson, B.L. and House, J.K. (2008) Serologic cross-reactivity of Australian,
- 383 Moraxella bovis to vaccinal bacterin strains as determined by competitive ELISA, Australian Veterinary
- 384 Journal, 86:4, pp. 124-129.
- 385 McDonnell, G. and Russell, A.D. (1999) Antiseptics and Disinfectants: Activity Action and Resistance, 12:1,
- 386 pp. 147-179.
- 387 Menendez, A.B., Parra, A.L., Pavon, V.B., Dominguez, C.C., Martinez, O.V. Sardinas, I.G. and Munoz, A.
- 388 (2007) Actividad Cicatrizante y Ensayos de Irritación de la Crema de Calendula officinalis al 1%, Lat. Am. J.
- 389 Pharm., 26:6, pp. 811-817.

July 12, 2017 Page 10 of 12

- 390 Mika, D. and Guruvayoorappan C. (2011) Myeloperoxidase: the yin and yang of tumour progression,
- Journal of Experimental Therapeutics and Oncology, 9, pp. 93-100.
- 392 Mika, Denish and C, Guruvayoorappan (2011) Myeloperoxidase: the vin and yang in tumor progression,
- Journal of Experimental Therapeutics and Oncology, 9, pp. 93-100.
- 394 Miles, C.M. and Wayne, M. (2008) Quantitative Trait Locus (QTL) Analysis, Scitable, Nature Education, 1,
- 395 pp. 1-4.
- 396 Moraxella bovis to vaccinal bacterin strains as determined by competitive ELISA Australian Veterinary
- 397 Journal, 86:4, pp.124-129.
- 398 Mouli, S.P. (2005) Copper sulphate in veterinary practice, Indian J. Vet Surg., 26:1, pp. 57-58.
- Nakae, H. and Inaba (2000) Effectiveness of Electrolyzed Oxidized Water Irrigation in a Burn-Wound
- 400 Infection Model, J Trauma, 49, pp. 511–514.
- 401 National Center for Biotechnology Information—NCBI (2017) Hypochlorous Acid, PubChem Compound
- Database; CID=24341, https://pubchem.ncbi.nlm.nih.gov/compound/24341 (accessed June 5, 2017).
- 403 O'Conner, A.M., Brace, S., Gould, S., Dewell, R. and Engelken, T. (2012) A randomized clinical trial
- 404 evaluating a farm-of-origin autogenous *Moraxella bovis* vaccine to control infectious bovine
- 405 keratoconjunctivis (Pinkeye) in beef cattle, J Vet Intern Med, 25, pp. 1447–1453.
- 406 OConnor, A.M., Brace, S., Gould, S., Dewell, R. and Engelken, T. (2011) A Randomized Clinical Trial
- 407 Evaluating a Farm-of-Origin Autogenous Moraxella bovis Vaccine to Control Infectious Bovine
- Keratoconjunctivis (Pinkeye) in Beef Cattle, J Vet Intern Med, 25, pp. 1447–1453.
- 409 OConnor, A.M., Shen, H.G., Wang, C. and Opriessnig, T. (2012) Descriptive epidemiology of Moraxella
- 410 bovis, Moraxella bovoculi and Moraxella ovis in beef calves with naturally occurring infectious bovine
- 411 keratoconjunctivitis (Pinkeye), Veterinary Microbiology, 155, pp. 374–380.
- OConnor, A.M., Shen, H.G., Wang, C. and Oriessnig, T. (2011) Descriptive epidemiology of Moraxella
- bovis, Moraxella bovoculi and Moraxella ovis in beef calves with naturally occurring infectious bovine
- 414 keratoconjunctivitis (Pinkeye), Veterinary Microbiology, 155, pp. 374–380.
- Pegrift, R.Y. and Friedman, A.J. (2013) Topical hypochlorous acid (HOCl) as a potential treatment of
- 416 pruritus, Curr Derm Rep, 2, pp. 181–190.
- 417 Pickens, L.G. and Miller, R.W. (1980) Biology and control of the face fly, Musca autumnalis
- 418 (Dipter:Muscidae), J. Med. Entomology, 17:3, pp. 195-210.
- Pullar, J.M., Vissers, M.C.M. and Winterbourn, C.C. (2000) Living with a Killer: The Effects of
- 420 Hypochlorous Acid on Mammalian Cells, Life, 50, pp. 259–266.
- Ramey, D.W. and Kinde, H. (2015) Commercial and Homemade Extremely Dilute Hypochlorous Acid
- 422 Solutions Are Bactericidal Against Staphylococcus aureus and Escherichia coli In Vitro, Journal of Equine
- 423 Veterinary Science, 35, pp. 161–164.
- 424 Robson, M.C., Payne, W.G., Ko, F.K., Mentis, M.Donati, G., Shafi, S.M., Culverhouse, S., Wang, L. Khosrovi,
- 425 B., Najafi, R., Cooper, D.M. and Bassiri, M. (2007) Hypochlorous Acid as a Potential Wound Care Agent,
- 426 Part II. Stabilized Hypochlorous Acid: Its Role in Decreasing Tissue Bacterial Bioburden and Overcoming
- 427 the Inhibition of Infection on Wound Healing, Journal of Burns and Wounds, 6, pp. 80-90.
- 428 Rosenberg, L., Lapid, O., Bogdanov-Berezovsky, Glesinger, R., Krieger, Y. Silberstein, E., Sagi, A. Judkins,
- 429 K. and Singer, A.J. (2004) Safety and efficacy of a proteolytic enzyme for enzymatic burn debridement: a
- 430 preliminary report, Burns, 30, pp. 843–850.
- 431 Sakarya, S., Gunay, N., Karakulak, M. Ozturk, B. and Ertugrul, B. (2014) Hypochlorous Acid: An Ideal
- Wound Care Agent With Powerful Microbicidal, Antibiofilm, and Wound Healing Potency, Wounds,
- 433 26:12, pp. 342-350.
- 434 Sakarya, S., Gunay, N., Meltem, K., Ozturk, B. and Ertugrul, B. (2016) Hypochlorous Acid: An ideal wound
- 435 care agent with powerful, antibiofilm, and wound healing potency, 26:12, pp. 342-350.

July 12, 2017 Page 11 of 12

- 436 Schneider, L.A., Korber, A., Grabbe, S. and Dissemond, J. (2007) Influence of pH on wound-healing: a new
- perspective for wound-therapy? Arch Dermatol Res, 298, pp. 413–420.
- 438 Selkon, J.B., Cherry, G.W., Wilson, J.M. and Hughes, M.A. (2006) Evaluation of hypochlorous acid washes
- in the treatment of chronic venous leg ulcers, Journal of Wound Care, 15:1, pp, 33-37.
- Skovgard, H. (2004) Sustained releases of the pupal parasitoid *Spalangia cameroni* (Hymenoptera:
- Pteromalidae) for control of house flies, *Musca domestica* and stable flies *Stomoxys calcitrans* (Diptera:
- 442 Muscidae) on dairy farms in Denmark, Biological Control, 30, pp. 288–297.
- Snowder, G.D., van Vleck, L.D., Cundiff, L.V. and Bennet, G.L. (2005) Genetic and environmental factors
- associated with incidence of infectious bovine keratoconjunctivitis in preweaned beef calves, Journal of
- 445 Animal Science, 83:3, pp. 507-518.
- 446 Steelman, C.D., Gbur, E.E., Tolley, G. and Brown, A.H. (1993) Variation in population density of the face
- fly, *Musca autmnalis* de Geer, among selected cattle breeds of beef cattle, J. Agric. Entomol., 10:2, pp. 97-106.
- Surface, H.A. (1915) To keep down house flies, Zool. Press bulletin, 313.
- 449 Swaim, S.F. and Lee, A.H. (1987) Topical Wound Medications: a review, Journal of the American
- 450 Veterinary Medical Association, 190:12, pp. 1588-1593.
- 451 Swaim, S.F. and Lee, A.H. (1987) Topical wound medications: a review, J. Amer. Veterinary Medical
- 452 Assoc., 12:15, pp. 1588-1593.
- 453 Takesue, Y., Takahashi, Y., Ichiki, K., Nakajima, K., Tsuchida, T. A., Uchino, M. and Ikeuchi, H. (2011)
- 454 Application of an Electrolyzed Strongly Acidic Aqueous Solution Before Wound Closure in Colorectal
- Surgery, Diseases of the Colon and Rectum, 54: 7, pp. 826-832.
- 456 US Department of Agriculture, National Organic Program US NOP (2015) Policy Memo 15-4
- Vijayaraghavan, K., Rajkumar, J., Bukhari, S.N.A., Al-Sayed, B. and Seyed, M.A. (2017) Chromolaena odorata:
- 458 A neglected weed with a wide spectrum of pharmacological activities (Review), Molecular Medicine
- 459 Reports, 15, pp. 1007-1016.
- 460 White, G. C. (1972) Handbook of Chlorination. Van Nostrand Reinhold Company, New York.
- Wiant, C. (2013) The chlorine residual: A public health safeguard, Water Quality and Health Council.
- 462 Zinke, J. (2010) Treatment of conjunctivitis and conjunctival irritation in animals, Biolgische tiermedizin,
- 463 27:1, pp. 17-25.

July 12, 2017 Page 12 of 12